

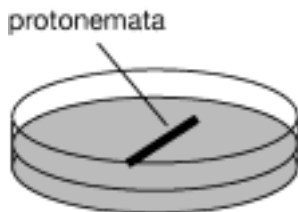
## Observation of mitosis in a protonemal cell of Physcomitrella

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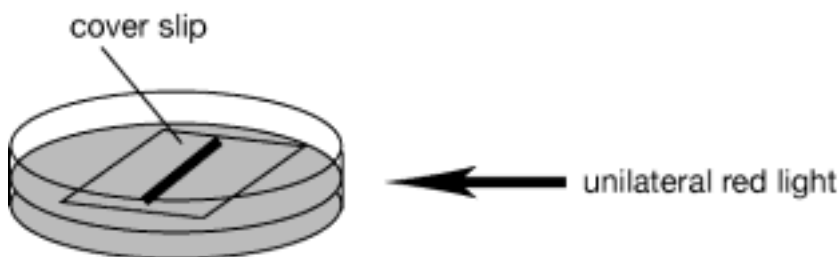
This protocol gives you the opportunity to see cell divisions of protonemal apical cells of Physcomitrella.

### Pre-culture

1. Sub-culture protonemal cells on cellophane-overlaid BCDATG\* plates under continuous white light ( $30\text{-}40\ \mu\text{mol m}^{-2}\ \text{sec}^{-1}$ ) at  $25^\circ\text{C}$  every 5~7 days.
2. Inoculate ~7-day-old protonemal cells into BCDATG plates (3 cm-diameter dish) and cover protonemal cells with a sterile cover slip (18 x18 mm).



3. Incubate the plates under unilateral red light ( $15\text{-}20\ \mu\text{mol m}^{-2}\ \text{sec}^{-1}$ ) at  $25^\circ\text{C}$  for 7 days. Red light can be obtained from fluorescent tube filtered through a red plastic sheet (We use acrylite 102; Mitsubishi Rayon, Japan, <http://www.mrc.co.jp/acrylite/acryindex.html>)



### \*BCDATG

1 stock solutions

solution A (x 100 )

Ca(NO <sub>3</sub> ) <sub>2</sub> ·4H <sub>2</sub> O	118 g
FeSO <sub>4</sub> ·7H <sub>2</sub> O	1.25 g

H <sub>2</sub> O	fill up to 1000 ml
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SolutionB (x 100 )

MgSO <sub>4</sub> ·7H <sub>2</sub> O	25 g
H <sub>2</sub> O	fill up to 1000 ml

Autoclaved

SolutionC (x 100 )

KH <sub>2</sub> PO <sub>4</sub>	25 g
	adjust pH to 6.5 with 4 M KOH
H <sub>2</sub> O	fill up to 1000 ml

Autoclaved

Solution D (x 100)

KNO <sub>3</sub>	101 g
FeSO <sub>4</sub> ·7H <sub>2</sub> O	1.25 g
H <sub>2</sub> O	fill up to 1000 ml

Alternative TES (x 1000)

CuSO <sub>4</sub> ·5H <sub>2</sub> O	55 mg
H <sub>3</sub> BO <sub>3</sub>	614 mg
CoCl <sub>2</sub> ·6H <sub>2</sub> O	55 mg
Na <sub>2</sub> MoO <sub>4</sub> ·2H <sub>2</sub> O	25 mg
ZnSO <sub>4</sub> ·7H <sub>2</sub> O	55 mg
MnCl <sub>2</sub> ·4H <sub>2</sub> O	389 mg
KI	28 mg
H <sub>2</sub> O	fill up to 1000 ml

Autoclaved

500mM Ammonium Tartrate (x 100 )

Ammonium Tartrate	92.05 g
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H2O	fill up to 1000 ml
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Autoclaved

50mM CaCl<sub>2</sub> (x 50 )

CaCl <sub>2</sub> ·2H <sub>2</sub> O	7.35 g
H2O	fill up to 1000 ml

Autoclaved

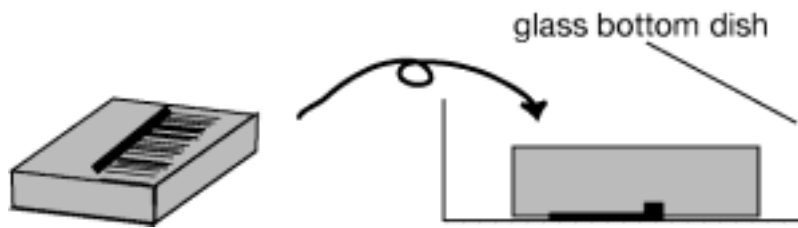
2.BCDATG

H2O	900 ml
Stock B	10 ml
Stock C	10 ml
Stock D	10 ml
Alternative TES	1 ml
500mM Ammonium Tartrate	10ml (= 5 mM)
50mM CaCl <sub>2</sub> · 2H <sub>2</sub> O (powder)	20 ml (= 1 mM) (0.15 g)
Glucose	5 g
Agar (Sigma, A6924)	8 g (= 0.8%)
	Fill up to 1000 mL with H2O

Autoclaved

### Time-lapse observation

1. Remove a cover slip from the plate and cut an agar block containing protonemal cell from the solid medium with a scalpel.
2. Turn the block up-side down and place it into 35 mm glass-bottom dish (IWAKI 3910-039: a 27 mm diameter opening in the center of a dish, [http://www.atgc.co.jp/div/rika/hbine/index\\_e.html](http://www.atgc.co.jp/div/rika/hbine/index_e.html)) as protonemal cell are touched on the bottom of the dish. Seal the dish with parafilm.



3. Place the dish on a stage of an inverted microscope. While you observe, room temperature should be set at 25°C.
4. Seek protonemal apical cell just before cell division and focus a nucleus. An apical cell before division is highly elongated and its cytoplasm is localized to apical side of a cell. One of the signs of mitosis is transition of a nuclear shape. Just before entering prophase, a nucleus will be a spherical shape rather than an oval shape.
5. Carry out the time-lapse observation. Mitosis will finish ~30 min in this condition. I usually acquire an image every 60 sec. If you examine the dynamics of GFP-fusion protein, you should not illuminate strong emission light to the cell because strong light not only fades GFP fluorescence but also inhibit cell division.